Title: Low smoking-exposure, the adolescent brain, and the modulating role of CHRNA5 polymorphisms

Authors: Bader Chaarani ^{1*}, Kees-Jan Kan ¹, Scott Mackey ¹, Philip A. Spechler ¹, Alexandra Potter ¹, Catherine Orr ¹, Nicholas D'Alberto ¹, Kelsey E. Hudson ¹, Tobias Banaschewski ², Arun L.W. Bokde ³, Uli Bromberg ⁴, Christian Büchel ⁴, Anna Cattrell ⁵, Patricia J.Conrod ^{6,7}, Sylvane Desrivières ⁵, Herta Flor ^{8,9}, Vincent Frouin ¹⁰, Jürgen Gallinat ¹¹, Penny Gowland ¹², Andreas Heinz ¹³, Bernd Ittermann ¹⁴, Jean-Luc Martinot ¹⁵, Frauke Nees ^{2,9}, Dimitri Papadopoulos-Orfanos ¹⁰, Tomáš Paus ¹⁶, Luise Poustka ^{2,17}, Michael N. Smolka ¹⁸, Henrik Walter ¹³, Robert Whelan ¹⁹, Stephen T. Higgins ¹, Gunter Schumann ⁶, Robert R. Althoff ¹, Elliot A. Stein ²⁰, Hugh Garavan ¹, & the IMAGEN Consortium ⁶.

Affiliations:

¹Vermont Center on Behavior and Health, Departments of Psychiatry and Psychology, University of Vermont, 05405 Burlington, Vermont, USA.

²Department of Child and Adolescent Psychiatry and Psychotherapy, Central Institute of Mental Health, Medical Faculty Mannheim, Heidelberg University, Square J5, 68159 Mannheim, Germany.

³Discipline of Psychiatry, School of Medicine and Trinity College Institute of Neurosciences, Trinity College Dublin, Ireland.

⁴University Medical Centre Hamburg-Eppendorf, House W34, 3.OG, Martinistr. 52, 20246, Hamburg, Germany.

⁵Medical Research Council - Social, Genetic and Developmental Psychiatry Centre, Institute of Psychiatry, Psychology & Neuroscience, King's College London, United Kingdom.

⁶Department of Psychiatry, Université de Montreal, CHU Ste Justine Hospital, Canada.

⁷Department of Psychological Medicine and Psychiatry, Institute of Psychiatry, Psychology & Neuroscience, King's College London, United Kingdom.

⁸Department of Cognitive and Clinical Neuroscience, Central Institute of Mental Health, Medical Faculty Mannheim, Heidelberg University, Square J5, Mannheim, Germany.

⁹Department of Psychology, School of Social Sciences, University of Mannheim, Mannheim, Germany.

¹⁰Neurospin, Commissariat à l'Energie Atomique, CEA-Saclay Center, Paris, France.

¹¹Department of Psychiatry and Psychotherapy, University Medical Center Hamburg-Eppendorf (UKE), Martinistrasse 52, 20246 Hamburg, Germany.

¹²Sir Peter Mansfield Imaging Centre School of Physics and Astronomy, University of Nottingham, University Park, Nottingham, United Kingdom.

¹³Department of Psychiatry and Psychotherapy, Campus Charité Mitte, Charité, Universitätsmedizin Berlin, Charitéplatz 1, Berlin, Germany.

¹⁴Physikalisch-Technische Bundesanstalt (PTB), Braunschweig and Berlin, Germany.

¹⁵Institut National de la Santé et de la Recherche Médicale, INSERM Unit 1000 "Neuroimaging & Psychiatry", University Paris Sud, University Paris Descartes - Sorbonne Paris Cité and Maison de Solenn, Paris, France.

¹⁶Rotman Research Institute, Baycrest and Departments of Psychology and Psychiatry, University of Toronto, Toronto, Ontario, M6A 2E1, Canada.

¹⁷Department of Child and Adolescent Psychiatry and Psychotherapy, Medical University of Vienna, Austria.

¹⁸Department of Psychiatry and Neuroimaging Center, Technische Universität Dresden, Dresden, Germany

¹⁹Department of Psychology, University College Dublin.

*Correspondence to:

Hugh Garavan UVM Department of Psychiatry hgaravan@uvm.edu

&

Bader Chaarani UVM Department of Psychiatry melmarsr@uvm.edu

Short title: Low smoking exposure and the adolescent brain.

Keywords: Neuroimaging; Low smoking exposure; Genetics; Grey matter volume; fMRI;

adolescents

Number of words in the abstract:206 Number of words in the text: 4078

Number of tables: 0. Number of figures: 4.

Number of supplementary material: 1.

²⁰ NIDA-IRP, NIH, Baltimore, USA.

Abstract:

Background: Studying the neural consequences of tobacco smoking during adolescence, including those associated with early light use, may inform the mechanisms that underlie the transition from initial use to nicotine dependence in adulthood. However, only a few studies in adolescents have appeared and those available include small samples. In addition, the neural mechanism, if one exists, that links nicotinic receptor genes to smoking behavior in adolescents is still unknown.

Methods: Structural and diffusion tensor magnetic resonance imaging data were acquired from a large sample of 14-year-old adolescents who completed an extensive battery of neuropsychological, clinical, personality and drug use assessments. Additional assessments were conducted at age 16.

Results: Exposure to smoking in adolescents, even at low doses, is linked to volume changes in the ventromedial prefrontal cortex and to altered neuronal connectivity in the corpus callosum. The longitudinal analyses strongly suggest that these effects are not pre-existing conditions in those who progress to smoking. There was a genetic contribution wherein the volume reduction effects were magnified in smokers who were carriers of the high-risk genotype of the alpha 5 nicotinic receptor subunit gene, rs16969968.

Conclusions: These findings give insight into a mechanism involving genes, brain structure and connectivity underlying why some adolescents find nicotine especially reinforcing.

Introduction

Smoking is the leading cause of preventable death in the United States and other developed countries. According to the U.S. Centers for Disease Control and Prevention (www.cdc.gov), smoking causes nearly six million deaths per year globally; current trends predict this will reach eight million by 2030. Using the U.S. as an example, more than five million Americans under 18 today are expected to die from a smoking-related illness, which is higher than the number of deaths caused by HIV, drug misuse, suicide, murder and motor vehicle injuries, combined. Smoking starts primarily during adolescence, with about 90% of US smokers reporting that they tried smoking before the age of 18. Every day in the US alone, 3,800 adolescents smoke their first cigarette and 2,100 become daily smokers. These statistics are particularly troubling given that early use of cigarettes during adolescence has been associated with heightened risk for later dependence (1,2). Remarkably, even relatively low rates of cigarette consumption during adolescence (e.g., two to four cigarettes per week) increase the risk of becoming nicotine dependent by early adulthood (1,3).

Adolescence is a period of considerable brain development (4,5) and it has been hypothesized that nicotine use during this critical period produces neurobiological changes that promote tobacco dependence later in life (6). Thus, studying the neural consequences of smoking during adolescence, including those associated with early use, may inform the mechanisms that underlie the transition from initial use to nicotine dependence in adulthood. In contrast to the extensive literature on adult smokers, few studies in adolescents have appeared and those available include small samples.

Nicotine dependence is highly heritable (7) and genome-wide association studies have

revealed a reliable association between dependence and single nucleotide polymorphisms (SNPs) at the 15q nicotinic acetylcholine receptor (nAChR) α 5- α 3- β 4 gene cluster (8–10). The most replicated SNPs associated with smoking and lung cancer are the α 3 subunit gene (CHRNA3) rs1051730 (11,12) and the α 5 subunit gene (CHRNA5) rs16969968 (8), which both convert aspartic acid (G allele) to asparagine (A allele), with A being the risk-allele. However, the neural mechanisms, if any exist, that link these SNPs to smoking are poorly understood. One possible mechanism linking the polymorphism to nicotine dependence is reduced dopamine-mediated reward processing due to A allele nAChR expression (9). For example, Sherva et al. concluded that the A allele was significantly related to enhanced pleasurable responses to a person's first cigarette (10). Hong et al. showed that rs16969968 is associated with a dorsal anterior cingulate–ventral striatal/extended amygdala circuit, in which the risk-allele was associated with decreased intrinsic resting functional connectivity strength in smokers and, to a lesser extent, nonsmokers. Further, the connectivity strength of the circuit distinguished smokers from nonsmokers and predicted addiction severity in smokers (13).

In the present study, we sought to determine how these genetic predispositions might impact the morphometry of the developing adolescent brain and its relationship to smoking. We were specifically interested in the genetic influence of the CHRNA3 SNP rs1051730 and the CHRNA5 SNP rs16969968 on initial cigarette use in order to illuminate possible pathways that lead to heavier use and dependence. Using multimodal neuroimaging in a large cohort of 14 years old adolescents from the IMAGEN study (http://www.imagen-europe.com), we first determined grey matter volume (GMV) differences in both early smokers and nonsmokers followed by the influence of the above two SNPs. Then, we measured white matter connectivity to determine the anatomical structural connectivity that may have supported these genetic polymorphism findings.

We hypothesize, based on previous neuroimaging studies, to find a smoking association with GMV mainly in the prefrontal cortex (14–16), and with white matter connectivity mainly in the corpus callosum (17–19). We also hypothesize, based on previous GWAS findings (8), to observe a small but significant association between CHRNA5 and smoking behavior.

Materials and Methods

Overview of IMAGEN protocols

Full details of the procedures employed by the IMAGEN study, including details on ethics, recruitment, standardized instructions for administration of the psychometric and cognitive behavioral measures are available in the standard operating procedures for the IMAGEN project (http://www.imagen-europe.com/en/Publications_and_SOP.php).

Participants

Data were acquired from 14-year-old adolescents. After complete description of the study to the participants and their parents/guardians, written informed consent was obtained. Individuals who provided assent completed an extensive battery of neuropsychological, clinical, personality and drug use assessments online and at the testing centers. Additional assessments were conducted at age 16. Participants were excluded if they had contraindications for MRI (metal or electronic implants and claustrophobia) or problematic medical history (e.g., diabetes, tumors, heart defects), neurological conditions (e.g., epilepsy, head trauma, neurodevelopmental disorders such as ADHD, OCD, depression and anxiety) or low IQ (<70). The Wechsler intelligence scale for children was used to measure IQ and was administered by experimenters at the study centers. The vocabulary and similarities subscales were employed to determine verbal IQ. The block design,

matrix reasoning and digit span subscales were employed to determine non-verbal/performance IQ. A puberty score was calculated using the Pubertal Development Scale (PDS) scale (20), which consisted of asking adolescents about physical development traits, such as growth in height, body hair, skin changes, and other gender-specific traits (i.e. voice deepening, menstruation). The PDS category scores (answers) are as follows: 1 (No), 2 (Yes (Barely)), 3 (Yes (Definitely)), 4 (Development Completed). Data were normed according to the subject's age.

Smoking score

A cigarette-smoking score was calculated for the 14 year old adolescents from the European School Survey Project on Alcohol and Drugs (ESPAD) (21) questionnaire, which asked: "On how many occasions (if any) during your lifetime have you smoked cigarettes?". The ESPAD category scores are as follows: Score (Lifetime occurrences): 0 (0), 1 (1-2), 2 (3-5), 3 (6-9), 4 (10-19), 5 (20-39), 6 (>40). Follow-up smoking scores at age 16 were also collected. To account for second-hand smoke exposure, parent smoking scores were calculated using the same ESPAD questionnaire. These scores were not significantly different between smokers and non-smokers in the sample $(1.6\pm0.5 \text{ vs } 1.3\pm0.8, \text{ respectively})$, with p=0.23. Similar data using the same scoring system were obtained for alcohol use where an alcohol score was calculated from a similar ESPAD questionnaire which asked: "On how many occasions (if any) during your lifetime have you had any alcoholic beverage to drink?".

Different numbers of smokers and non-smokers were available for the neuroimaging and genetic analyses, which is graphically presented in Figure 1 and subject demographics detailed in supplementary Table 1.

Neuroimaging

MRI acquisition

MRI scanning was performed at the eight IMAGEN assessment sites (London, Nottingham, Dublin, Mannheim, Dresden, Berlin, Hamburg, and Paris) with 3T whole body MRI systems made by several manufacturers (Siemens: 4 sites, Philips: 2 sites, General Electric: 1 site, and Bruker: 1 site). To ensure comparability of MRI data acquired on these different scanners, image-acquisition techniques were implemented using a set of parameters compatible with all scanners that were held constant across sites, for example, those directly affecting image contrast or fMRI preprocessing. The full details of the MRI acquisition protocols and quality checks have been described previously, including the extensive period of standardization across MRI scanners (22).

Structural MRI

High-resolution anatomical MRIs were acquired with a three-dimensional T1 weighted magnetization prepared gradient echo sequence (MPRAGE) based on the ADNI protocol (http://www.loni.ucla.edu/ADNI/Cores/index.shtml).

Diffusion MRI

Diffusion weighted images were acquired with a single-shot EPI sequence with a b value of 1300 s/mm², a TE of 104 ms and a voxel size of 2.4x2.4x2.4mm, with 60 slices providing whole-brain coverage.

MRI data preprocessing

Preprocessing of the structural T1-weighted data was performed centrally with the Statistical Parametric Mapping version 8 (Wellcome Department of Neuroimaging, London, United Kingdom, http://www.fil.ion.ucl.ac.uk/spm/software/spm8/), using standard automated pipelines (22). Structural T1-weighted MRI processing included image segmentation into grey matter (GM), white matter and cerebrospinal fluid tissue classes, preceded by an iterative registration to the Montreal Neurological Institute (MNI) template space, using SPM's optimized normalization routine (23). For voxel-based morphometry (VBM), gray matter images were smoothed with a Full Width at Half Maximum (FWHM) Gaussian kernel of 8 mm, warped to standard MNI space and modulated by multiplying the linear and non-linear component of the Jacobian determinants generated during spatial normalization. Thus, the dependent measure in subsequent statistical analyses was absolute grey matter volume (GMV), facilitating comparisons of volumetric, rather than tissue concentration differences (24).

The Diffusion Tensor Imaging (DTI) preprocessing was performed with FSL 4.1 (www.fmrib.ox.ac.uk/fsl). The pipeline consisted of the following steps: First, Eddy-current and motion correction were applied using an affine registration and the B0 volume of the DTI data as reference. After registration, a brain extraction was applied to remove non-brain tissues and a first estimation of the diffusion tensor was achieved for each voxel. After B0 unwarping using the magnitude and phase images from the fieldmap acquisition, a second estimation of the diffusion tensor was achieved for each voxel. Finally, Fractional Anisotropy (FA) maps were generated for each subject.

Genetics

Genotyping and Quality control

DNA purification and genotyping was performed by the Centre National de Génotypage in Paris. DNA was extracted from whole blood samples preserved in ethylene-eiamine-tetra-acetic acid vacutainer tubes (BD, Becton, Dickinson and Company, Oxford, United Kingdom) using Gentra Puregene Blood Kit (QIAGEN, Valencia, California) according to the manufacturer's instructions. Genotype information was collected at 582,892 markers using the Illumina HumanHap610 and HumanHap660 Genotyping BeadChips (San Diego, California). The SNPs with call rates of <95%, minor allele frequency <1%, deviation from the Hardy-Weinberg equilibrium ($p \le 1E-06$), and non-autosomal SNPs were excluded.

Imputation of markers data

Markers data imputation and quality control for ambiguous SNPs, low MAF, missingness and HWE were done with MACH (25) following the ENIGMA2 guidelines (26). The 1000 Genomes project reference set of markers (http://www.1000genomes.org/data) was used for the imputation after decreasing the markers from ~41 million to ~13 million relevant genetic variants observed more than once in the European populations (26). Both rs16969968 and rs1051730 data were imputed and had good imputation quality (R^2 imputation quality metric ≥ 0.87) (supplementary Table 2).

Missing demographic data

Participants with missing data on sex and/or site were excluded. Missing values on continuous variables were replaced with the mean derived according to the participant's site and

sex. Missing values on nominal data were replaced with the mode of that variable for the participant's site and sex. The maximal missing rate for each variable was lower than 10%.

Statistical Analyses

Single SNP association analysis

Genotype effects of rs16969968 and rs1051730 on smoking levels were examined using single SNP linear regression in the 1417 subjects included in the genetic analysis (described in supplementary Fig 1-c and supplementary Table 1-Genetic analysis). The frequencies of the highrisk, the intermediate risk and the normal (no risk) genotypes for the two SNPs are described in supplementary Table 2. Four multidimensional scaling components were calculated using a metric model to account for population stratification and were included, in addition to age and gender, as covariates in an additive regression model (genotypes coded as 0, 1, and 2 for the number of risk alleles). These analyses performed using **PLINK** were v1.9 (http://pngu.mgh.harvard.edu/~purcell/plink/). No other SNPs were investigated in this study.

Brain voxel-wise analyses

Whole brain voxel-wise multiple regression analyses were performed on GMV and FA maps to identify regions significantly correlated with the smoking score and test whether effects were observable across the full range of smoking exposure levels (i.e. scores 0-6). These analyses were carried out using the general linear model, performed with the VBM toolbox of SPM8. Age, sex, PDS, handedness, scanner site (dummy coded), Performance IQ, Verbal IQ, socio-economic status and total GMV (only for GMV analyses) were included as nuisance covariates in the design matrix in all analyses. Performance IQ was significantly lower and alcohol use significantly higher

in smokers (p 's<0.05) and were included in the subsequent ROI-level analyses. The other variables were not different between the two groups (p 's>0.1).

The resulting set of voxel values constituted a Statistical Parametric Map of the t statistic (SPM $\{t\}$). We used 3dClustSim, a cluster correction Monte Carlo procedure available in AFNI (http://afni.nimh.nih.gov/), to determine a minimum cluster size that achieves a corrected significance of p < 0.05 with a voxel-wise threshold of p < 0.001 and a Full Width at Half Maximum (FWHM) spatial blur that is empirically derived from the spatial autocorrelation in the datasets (residuals from the voxelwise statistical analyses). Clusters with a spatial extent threshold > 411 voxels were considered significantly related to smoking levels.

ROI-level analysis

The mean GMV was extracted with the Marsbar toolbox (27) from the ROI that the VBM regression analysis revealed to be significantly associated with smoking. GMV values were then included as a dependent variable in a 2 (smoking status) by 3 (genotype) ANCOVA model to test the interaction between smoking and genotype on GMV. In this model, subjects were grouped into smokers (score = 1-6) and non-smokers (score = 0) then into the three genotypes (AA, GA and GG). Further, the mean FA was extracted from the ROI obtained from the DTI regression analysis that was significantly associated with smoking and then included as a dependent variable in a similar ANCOVA model to test the interaction between smoking amount and genotype on FA.

Results

Smoking status and CHRNA genotype effects on structural variations in the cortex

First, we studied the neuroanatomical correlates of cigarette use in grey matter density in 211 adolescent smokers from the cohort who had VBM data passing quality control and very low to moderate smoking exposure (smoking scores 1-6), compared with 627 non-smokers (Figure 1-A). A whole brain VBM regression analysis showed a significant negative linear relation between GMV and smoking scores (r = -0.2, $p = 5E^{-06}$, df=822) in the ventromedial prefrontal cortex (vmPFC) (cluster size: 499 voxels; cluster peak coordinates: x = 6, y = 30, z = -12) after p < 0.05 cluster-wise correction (Figure 2-A).

We next replicated the association of the rs16969968 genotype previously reported in smokers (9) in 940 non-smokers and 477 smokers across all smoking ranges (Figure 1-C) (p=0.003), which explained ~ 3.5% of smoking behavior variance in the sample (Supplementary Table 2). Analyses of the rs1051730 SNP revealed a similar result to that seen for rs16969968, which is not surprising given their strong linkage disequilibrium (D'=1; R² = 0.99; supplementary Table 2). Assuming that rs16969968 is the functional locus (9), results from rs1051730 are not reported further and subsequent analyses are restricted to rs16969968.

Next, we studied smoking exposure * genotype interaction effects on the lower vmPFC volume derived from the whole brain regression analysis that provided the vmPFC ROI used for subsequent analyses. A 2 (smoking status) by 3 (genotype) ANCOVA indicated that smoking status (all smoking-exposure levels; p < 0.0005) and the smoking*genotype interaction (p = 0.02) had significant effects on the vmPFC volume (Figure 2-C); there was no significant main effect for genotype in either the whole sample or in non-smokers (p > 0.09). Notably, the ANCOVA showed that the vmPFC volume was significantly lower in smokers in each of the three genotypes,

compared with non-smokers (p = 0.000012), with the largest effect in homozygote carriers of the high-risk alleles (AA genotype; n= 18 smokers and n = 58 non-smokers). Moreover, smoking levels did not differ significantly among the three genotype groups (p=0.1).

Alcohol use and Performance IQ effects in smokers

In our sample, alcohol use and performance IQ were significantly associated with smoking (supplementary Table 1), thus rendering it difficult to attribute the vmPFC anatomical and functional effects to smoking *per se*. To address this, we identified 341 non-smokers and assessed the correlations between alcohol use (alcohol scores ranging from 0-6 reflecting the same ranges of lifetime use as defined for smoking) and performance IQ with the volumetric measures for the vmPFC. The same covariates were included as above. The results yielded no significant associations with alcohol use ($R^2 < 0.0005$, p > 0.1, df=326) or performance IQ ($R^2 < 0.01$, p > 0.05, df=325) (Figure 3). These results strongly suggest that alcohol use and performance IQ, on their own (i.e., in non-smokers), do not significantly impact the volumetric effects in the vmPFC and thus are unlikely to be the source of the observed genetic effects in smokers.

Smoking status effects on white matter

Finally, we examined the connectivity of the white matter as a function of smoking. Using the same covariates as in the previous analyses (except for total GMV), we performed a whole-brain regression on a sample of 147 smokers and 529 non-smokers for a total of 676 subjects (Figure 1-B). The analysis yielded a significant negative correlation (r=-0.24, $p = 5E^{-06}$, df=661) between FA and smoking scores, in the anterior corpus callosum (359 voxels, x=-15, y=30, z=10)

after p corrected<0.05 (Figure 4-A), reflecting altered inter-hemispheric axonal structural properties with smoking and revealing a linear relationship similar to what was observed between the vmPFC volume and smoking. Finally, a 2 X 3 ANCOVA revealed no significant interaction between smoking and genotype on FA extracted from the corpus callosum ROI (p>0.1). While this could be explained by the lack of statistical power due to the loss of a significant number of heavier smokers who did not have DTI data, it is also possible that the nicotinic receptor genetic influence was manifest on GMV but not white matter connectivity.

Discussion

In this study, we have shown that exposure to tobacco smoking in adolescents, even at low doses, is linked to a reduction in vmPFC gray matter volume and altered neuronal connectivity in the corpus callosum. Most notably, the regression analyses indicate linear reductions in vmPFC volumes and neuronal connectivity observable in the very light smoking group of young adolescents. Finally, there was a small yet interesting genetic contribution wherein the vmPFC volume reduction effects were magnified in smokers who were carriers of the high-risk polymorphisms of the alpha 5/alpha 3 nicotinic receptor subunits. The absence of both main effects of genotype and any genotype effects in non-smokers indicates a gene * exposure interaction such that the effects of the polymorphisms are only evident if the adolescent is a smoker.

Our structural findings are in line with numerous VBM studies reporting negative doseresponse correlations between the PFC volume/density in general (14–16) or the vmPFC volume in particular (28) and lifetime cigarette usage in adult heavy smokers. Notably, the present results show this relationship in a group of relatively inexperienced adolescent smokers and suggests the linear decrease is present even at the lightest levels of smoking. These results support a growing literature suggesting that smoking particularly affects the PFC, either from nicotine or one of the more than four thousand chemicals present in tobacco, about 400 of which, including nicotine and CO, are known toxins (29).

Despite the converging evidence of apparent brain atrophy in moderate and heavy adult smokers in the PFC (14–16), the dose-response relationship observed in our data is intriguing as it suggests that just one or two cigarettes can potentially alter adolescent cortex development, an observation that has not been previously reported. An alternative interpretation is that the lower vmPFC GMV preceded and predisposed toward smoking initiation. To address this possibility, we assessed the relationship between vmPFC volume at age 14 and future smoking at age 16 in 627 adolescents who were smoking naïve at age 14, using the same covariates included in the previous regression analyses. Within this sample of adolescents, 386 remained never-smokers at age 16 with the remaining 241 showing a similar use distribution in smoking levels at age 16 as was observed in the previous analysis of smokers at age 14 (see Figure 1). The vmPFC volume derived from the age 14 regression analysis did not predict future smoking at age 16 ($\beta = 0.02$, p = 0.9, df=611) (Figure 2-B), which does not support the hypothesis that the observed volume reduction predisposed towards adolescent use. Rather, this finding is consistent with the interpretation that even extremely low smoking exposure by age 14 may influence brain maturation in early adolescence.

The association of the rs16969968 genotype with smoking behavior is consistent with Hong et al (30) who showed that rs16969968 genotype significantly explains 3.3% and 4.6% of the variance of nicotine addiction severity and cigarettes per day, respectively. The gene*exposure vmPFC effect is consistent with a recent meta-analysis (31) of pharmacological neuroimaging studies which revealed that cigarette smoking and CHRNA agonist administration in adult smokers

are both associated with lower neural activity in, among other regions, the vmPFC. In line with these findings, another meta-analysis, interrogating the neurobiological targets of pharmacological and cognitive-based treatments for addiction to nicotine, identified similar portions of the vmPFC to have lower activity in smokers (32). Moreover, the rs16969968 * smoking interaction provides evidence that nicotine, rather than the other chemicals in cigarettes, might be the basis of the association between smoking and GMV reductions modulated by the nicotinic acetylcholine receptor system.

The DTI findings are in line with previous data showing that smokers have lower white matter FA in the anterior corpus callosum (17–19), which has been interpreted to indicate possible axonal damage and disrupted myelin integrity in the region (19). Conversely, other studies have reported higher (33) or unchanged FA (34) in the corpus callosum of smokers. This discordance could be explained by the fact that the used sample differed by age, size and/or smoking exposure. Similar to the GMV findings, FA differences were observed even in very light smokers. To address the possibility that the reduced FA values, like the reduced vmPFC GMV, preceded smoking, we similarly assessed the relationship between anterior corpus callosum FA at age 14 and future smoking at age 16 in 531 adolescents who were smoking naïve at age 14 (348 never-smokers and 183 smokers), using the same covariates as in the previous FA analyses. The anterior corpus callosum FA, derived from the regression analysis, did not predict future smoking (r=0.02, p=0.89, df=516) (Figure 4-B), which supports the conclusion that the FA reduction did not precede adolescent use but rather that very low smoking exposure appears to alter adolescent brain development.

Since the anterior corpus callosum connects regions of the prefrontal cortex with morphologically similar regions in the opposite hemisphere (35), we next asked if the anterior corpus callosum FA reduction was related to the vmPFC volume reduction of the age 14 smokers. There was a significant positive correlation between the two in smokers and non-smokers (r=0.25, p<0.0001, df=615; Figure 4-C) within 630 adolescents (498 non-smokers and 132 smokers) from the sample used for the whole brain FA regression analysis who also had GMV data. Finally, while the absence of interaction between smoking and genotype on FA could be explained by the lack of statistical power due to the loss of a significant number of heavier smokers who did not have DTI data, it is also possible that the nicotinic receptor genetic influence was manifest on grey matter volume but not white matter connectivity.

The key limitation of the present study is the sample size for the genetic analyses. With 1417 participants on whom genetic data was available (940 non-smokers, 417 smokers), we are at the lower limit of the genetic-association approaches for estimating contributions of common SNPs to phenotypic variations, especially because individual SNPs only explain small amounts of this variance. The limited sample size also affected the significance values in the brain-genotype interaction analysis where we ended up with a small group (N=18) of smokers having the double risk allele. The genetic findings must be therefore interpreted in light of the sample size limitations. Nevertheless, this sample size limitation must be viewed in the context of the phenotype under study. In fact, the IMAGEN dataset is currently the largest longitudinal brain imaging and genetics study in adolescents worldwide, allowing us to detect unique relationships, even if small, between the brain, genetics and smoking behavior.

Combined, this study's results indicate a structural and functional basis for dose-response changes in the brain of young adolescent smokers, which may underlie, at least in part, the known CHRNA genetic association with smoking. The longitudinal analyses suggest that these effects are not pre-existing conditions in those who progress to smoking, but rather may be, in the case of

GMV for example, an initial phase of volume reductions of the PFC in general and the vmPFC in particular that have been observed in adult heavy smokers (14–16,28). Although adolescent experimentation with smoking is common, these results give insight into a mechanism involving genes, brain structure and brain connectivity underlying why some teens find nicotine especially reinforcing and transition to repeated use leading to increased risk of lifetime dependence.

Acknowledgments:

This work received support from the following sources: P50DA036114 from the National Institute on Drug Abuse and Food and Drug Administration; P20GM103644 from the National Institute on General Medical Sciences; the European Union-funded FP6 Integrated Project IMAGEN (Reinforcement-related behaviour in normal brain function and psychopathology) (LSHM-CT- 2007-037286), the FP7 projects IMAGEMEND (602450; IMAging GEnetics for MENtal Disorders) and MATRICS (603016), the Innovative Medicine Initiative Project EU-AIMS (115300-2), a Medical Research Council Programme Grant "Developmental pathways into adolescent substance abuse" (93558), the Swedish funding agency FORMAS, the Medical Research Council and the Wellcome Trust(Behavioural and Clinical Neuroscience Institute, University of Cambridge), the National Institute for Health Research (NIHR) Biomedical Research Centre at SouthLondon and Maudsley NHS Foundation Trust and King's College London, the Bundesministerium fur Bildung und Forschung (BMBF grants 01GS08152; 01EV0711; eMED SysAlc 01ZX1311A; Forschungsnetz AERIAL), the Deutsche Forschungsgemeinschaft (DFG): Reinhart-Koselleck Award SP 383/5-1 and grants SM 80/7-1, SFB 940/1, FOR 1617), the French MILDT (Mission Interministe rielle de Lutte contre la Drogue et la Toxicomanie), the CENIR (Centre de NeuroImagerie de Recherche, Pr. S. Lehe ricy) within the ICM institute, the National Institute of Mental Health (MH082116), and a National Institutes of Health Center of Biomedical Research Excellence award P20GM103644 from the National Institute of General Medical Sciences and the Tobacco Centers of Regulatory Science award P50DA036114. HG was supported, in part, by NIH Consortium grant U54 EB020403, supported by a cross-NIH alliance that funds Big Data to Knowledge Centers of Excellence.

G.S. is the primary investigator of the IMAGEN Consortium. T.B., G.B, A.L.W.B., U. B., C.B., A.C., P.J.C., S.D., H.F., V.F., J.G., P.G., A.H., B.I., J.-L.M., M-L.P-M., F.N., D.P.O.,L. P., T.P., M.N.S., H.W., R.W., G.S. and H.G acquired and pre-processed the data. B.C. and H.G. carried out the analyses and wrote the manuscript. K.J.K., S.M., P.S., N.D., K.E.H. & C.O. assisted in analytic methods. E.A.S., S.T.H., A.P., & R.R.A. provided expert advice and edited the manuscript.

Members of the IMAGEN Consortium not listed as individual authors include: Karl Mann, Maren Struve, Marcella Rietschel, Rainer Spanagel, Mira Fauth-Bühler, Sabina Millenet, and Yvonne Grimmer at the Central Institute of Mental Health, University of Heidelberg; Nikolay Ivanov, Nicole Strache, Michael Rapp, Andreas Ströhle, and Jan Reuter at Charité, Universitätsmedizin Berlin; Alexis Barbot, Benjamin Thyreau, Yannick Schwartz, and Christophe Lalanne at the Comissariat à l'Energie Atomique; Jean-Luc Martinot, Zuleima Bricaud, Fanny Gollier Briand, Hervé Lemaitre, Jessica Massicotte, Helene Vulser, Jani Pentillä, and André Galinowski at the Institut national de la santé et de la recherche médicale; Tianye Jia, Helen Werts, Lauren Topper, Laurence Reed, Chris Andrew, Catherine Mallik, Barbara Ruggeri, Charlotte Nymberg, Lindsay Smith, Eva Loth, Stephanie Havatzias, Kerstin Stueber, and Argyris Stringaris at the Institute of Psychiatry, King's College London; Patrick Constant at PERTIMM (AsnièresSur-Seine); Ruediger Brühl, Albrecht Ihlenfeld, and Bernadeta Walaszek at the PhysikalischTechnische Bundesanstalt; Thomas Hübner, Kathrin Müller, Stephan Ripke, Sarah Rodehacke, Eva Mennigen, Dirk Schmidt, NoraVetter, and Veronika Ziesch at the Technische Universität Dresden; Jennifer Jones at the University College Dublin; Jean-Baptiste Poline at the University of California, Berkeley; Tahmine Fadai, Juliana Yacubian, and Sophia Schneider at the University of Hamburg; Claire Lawrence, Craig Newman, Kay Head, and Nadja Heym at the University of Nottingham; and Zdenka Pausova, and Amir Tahmasebi at the University of Toronto.

Conflict of interest

The authors declare that there are not any competing financial interests in relation to the work described.

References:

- 1. Lydon DM, Wilson SJ, Child A, Geier CF. Adolescent Brain Maturation and Smoking: What We Know and Where We're Headed. Neurosci Biobehav Rev. 2014 Sep;45:323–42.
- 2. Kendler KS, Myers J, Damaj MI, Chen X. Early smoking onset and risk for subsequent nicotine dependence: a monozygotic co-twin control study. Am J Psychiatry. 2013 Apr;170(4):408–13.
- 3. Riggs NR, Chou C-P, Li C, Pentz MA. Adolescent to emerging adulthood smoking trajectories: when do smoking trajectories diverge, and do they predict early adulthood nicotine dependence? Nicotine Tob Res Off J Soc Res Nicotine Tob. 2007 Nov;9(11):1147–54.
- 4. Paus T, Keshavan M, Giedd JN. Why do many psychiatric disorders emerge during adolescence? Nat Rev Neurosci. 2008 Dec;9(12):947–57.
- 5. Miguel-Hidalgo JJ. Brain structural and functional changes in adolescents with psychiatric disorders. Int J Adolesc Med Health. 2013;25(3):245–56.
- 6. DeBry SC, Tiffany ST. Tobacco-induced neurotoxicity of adolescent cognitive development (TINACD): a proposed model for the development of impulsivity in nicotine dependence. Nicotine Tob Res Off J Soc Res Nicotine Tob. 2008 Jan;10(1):11–25.
- 7. Lessov-Schlaggar CN, Pergadia ML, Khroyan TV, Swan GE. Genetics of nicotine dependence and pharmacotherapy. Biochem Pharmacol. 2008 Jan 1;75(1):178–95.
- 8. Bierut LJ. Convergence of genetic findings for nicotine dependence and smoking related diseases with chromosome 15q24-25. Trends Pharmacol Sci. 2010 Jan;31(1):46–51.
- 9. Bierut LJ, Stitzel JA, Wang JC, Hinrichs AL, Grucza RA, Xuei X, et al. Variants in nicotinic receptors and risk for nicotine dependence. Am J Psychiatry. 2008 Sep;165(9):1163–71.
- 10. Sherva R, Wilhelmsen K, Pomerleau CS, Chasse SA, Rice JP, Snedecor SM, et al. Association of a single nucleotide polymorphism in neuronal acetylcholine receptor subunit alpha 5 (CHRNA5) with smoking status and with "pleasurable buzz" during early experimentation with smoking. Addict Abingdon Engl. 2008 Sep;103(9):1544–52.
- 11. Leung T, Bergen A, Munafò MR, De Ruyck K, Selby P, De Luca V. Effect of the rs1051730–rs16969968 variant and smoking cessation treatment: a meta-analysis. Pharmacogenomics. 2015 May 1;16(7):713–20.
- 12. Munafò MR, Johnstone EC, Walther D, Uhl GR, Murphy MFG, Aveyard P. CHRNA3 rs1051730 genotype and short-term smoking cessation. Nicotine Tob Res Off J Soc Res Nicotine Tob. 2011 Oct;13(10):982–8.

- 13. Hong LE, Hodgkinson CA, Yang Y, Sampath H, Ross TJ, Buchholz B, et al. A genetically modulated, intrinsic cingulate circuit supports human nicotine addiction. Proc Natl Acad Sci. 2010 Jul 27;107(30):13509–14.
- 14. Brody AL, Mandelkern MA, Jarvik ME, Lee GS, Smith EC, Huang JC, et al. Differences between smokers and nonsmokers in regional gray matter volumes and densities. Biol Psychiatry. 2004 Jan 1;55(1):77–84.
- 15. Gallinat J, Meisenzahl E, Jacobsen LK, Kalus P, Bierbrauer J, Kienast T, et al. Smoking and structural brain deficits: a volumetric MR investigation. Eur J Neurosci. 2006 Sep 1;24(6):1744–50.
- 16. Zhang X, Salmeron BJ, Ross TJ, Geng X, Yang Y, Stein EA. Factors underlying prefrontal and insula structural alterations in smokers. NeuroImage. 2011 Jan 1;54(1):42–8.
- 17. Lin F, Wu G, Zhu L, Lei H. Heavy smokers show abnormal microstructural integrity in the anterior corpus callosum: a diffusion tensor imaging study with tract-based spatial statistics. Drug Alcohol Depend. 2013 Apr 1;129(1–2):82–7.
- 18. Savjani RR, Velasquez KM, Thompson-Lake DGY, Baldwin PR, Eagleman DM, De La Garza R, et al. Characterizing white matter changes in cigarette smokers via diffusion tensor imaging. Drug Alcohol Depend. 2014 Dec 1;145:134–42.
- 19. Viswanath H, Velasquez KM, Thompson-Lake DGY, Savjani R, Carter AQ, Eagleman D, et al. Alterations in interhemispheric functional and anatomical connectivity are associated with tobacco smoking in humans. Front Hum Neurosci [Internet]. 2015 Mar 9 [cited 2016 Feb 3];9. Available from: http://www.ncbi.nlm.nih.gov/pmc/articles/PMC4353249/
- 20. Petersen AC, Crockett L, Richards M, Boxer A. A self-report measure of pubertal status: Reliability, validity, and initial norms. J Youth Adolesc. 1988 Apr;17(2):117–33.
- 21. Hibell B. The 1995 ESPAD report: alcohol and other drug use among students in 26 European countries. European School Survey Project on Alcohol and other Drugs, editor. 1997;
- 22. Schumann G, Loth E, Banaschewski T, Barbot A, Barker G, Büchel C, et al. The IMAGEN study: reinforcement-related behaviour in normal brain function and psychopathology. Mol Psychiatry. 2010 Dec;15(12):1128–39.
- 23. Ashburner J, Friston KJ. Unified segmentation. NeuroImage. 2005 Jul 1;26(3):839–51.
- 24. Ashburner J, Friston KJ. Voxel-Based Morphometry—The Methods. NeuroImage. 2000 Jun;11(6):805–21.
- 25. Li Y, Willer CJ, Ding J, Scheet P, Abecasis GR. MaCH: using sequence and genotype data to estimate haplotypes and unobserved genotypes. Genet Epidemiol. 2010 Dec;34(8):816–34.

- 26. ENIGMA2 Genetics Support Team. ENIGMA2 1KGP Cookbook (v3) [Online]. The Enhancing Neuroimaging Genetics trhough Meta-Analysis (ENIGMA) Consortium. [Internet]. 2013. Available from: http://enigma.ini.usc.edu/wp-content/uploads/2012/07/ENIGMA2_1KGP_cookbook_v3.pdf
- 27. Brett M. Region of interest analysis using an SPM toolbox [abstract]. Present 8th Int Conferance Funct Mapp Hum Brain. 2002 Jun 2;16(2):abstract 497.
- 28. Fritz H-C, Wittfeld K, Schmidt CO, Domin M, Grabe HJ, Hegenscheid K, et al. Current Smoking and Reduced Gray Matter Volume—a Voxel-Based Morphometry Study. Neuropsychopharmacology. 2014 Oct;39(11):2594–600.
- 29. Swan GE, Lessov-Schlaggar CN. The effects of tobacco smoke and nicotine on cognition and the brain. Neuropsychol Rev. 2007 Sep;17(3):259–73.
- 30. Hong LE, Yang X, Wonodi I, Hodgkinson CA, Goldman D, Stine OC, et al. A CHRNA5 Allele Related to Nicotine Addiction and Schizophrenia. Genes Brain Behav. 2011 Jul;10(5):530–5.
- 31. Sutherland MT, Ray KL, Riedel MC, Yanes JA, Stein EA, Laird AR. Neurobiological Impact of Nicotinic Acetylcholine Receptor Agonists: An Activation Likelihood Estimation Meta-Analysis of Pharmacologic Neuroimaging Studies. Biol Psychiatry. 2015 Nov 15;78(10):711–20.
- 32. Konova AB, Moeller SJ, Goldstein RZ. Common and distinct neural targets of treatment: changing brain function in substance addiction. Neurosci Biobehav Rev. 2013 Dec;37(10 Pt 2):2806–17.
- 33. Hudkins M, O'Neill J, Tobias MC, Bartzokis G, London ED. Cigarette smoking and white matter microstructure. Psychopharmacology (Berl). 2012 Jan 4;221(2):285–95.
- 34. Paul RH, Grieve SM, Niaura R, David SP, Laidlaw DH, Cohen R, et al. Chronic cigarette smoking and the microstructural integrity of white matter in healthy adults: a diffusion tensor imaging study. Nicotine Tob Res Off J Soc Res Nicotine Tob. 2008 Jan;10(1):137–47.
- 35. Park H-J, Kim JJ, Lee S-K, Seok JH, Chun J, Kim DI, et al. Corpus callosal connection mapping using cortical gray matter parcellation and DT-MRI. Hum Brain Mapp. 2008 May;29(5):503–16.
- 41. Eklund, Anders, Thomas E. Nichols, and Hans Knutsson. "Cluster Failure: Why fMRI Inferences for Spatial Extent Have Inflated False-Positive Rates." *Proceedings of the National Academy of Sciences* 113, no. 28 (July 12, 2016): 7900–7905.

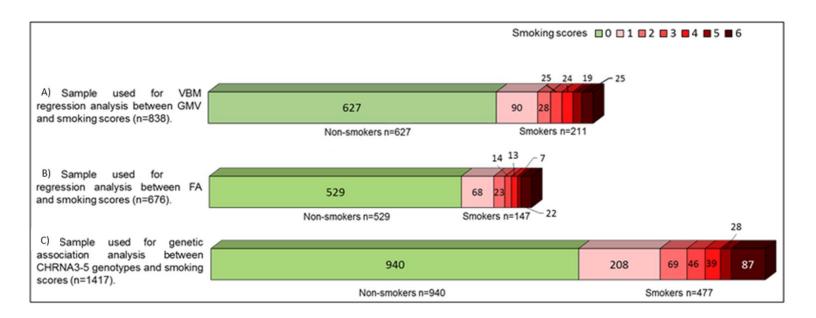
Figure Legends

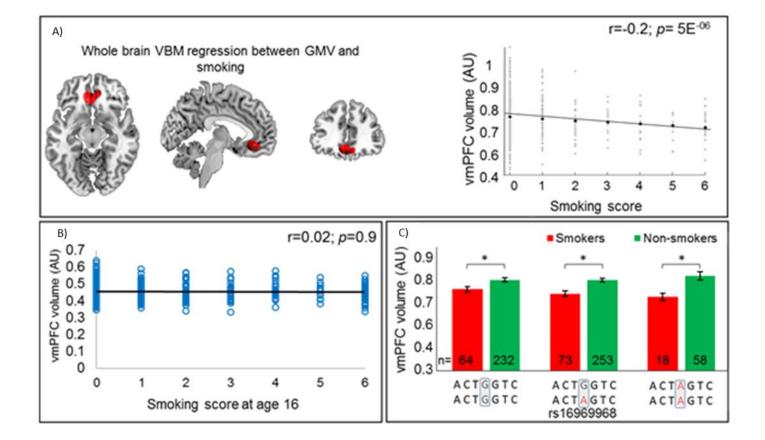
Figure 1. The different participant groups available from the IMAGEN study and used for the neuroimaging regression analyses (A, B) as well as for the genetic analyses (C). Non-smokers are represented with green bars (smoking score = 0). Smokers included in regression and genetic analyses had very low, low and moderate smoking exposure levels (smoking scores 1 to 6). Numbers on bars represent the total number of participants for each smoking score.

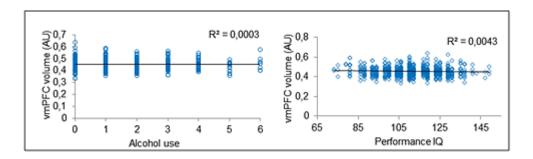
Figure 2. A) Whole brain rendering of the T maps resulting from the brain voxel-wise regression analysis between GMV and smoking score. A significant negative correlation (initial threshold p<0.005; p<0.05 when corrected for multiple comparisons) was observed in the ventromedial prefrontal cortex (vmPFC). No positive correlations between GMV and smoking score were detected. B) The relationship between the vmPFC volume and future smoking at age 16 in 627 adolescents who were smoking naïve at age 14. No significant correlation was observed with r=0.02 and p=0.9. C) The rs16969968 genotype effects on structural GMV in the vmPFC. A 2x3 ANCOVA indicated that smoking status and the smoking-genotype interaction had significant effects on the vmPFC volume (p<0.0005 and p=0.026, respectively), where it was significantly decreased in smokers with the effect being largest in the carriers of the smoking-related high-risk genotype (AA). (*): significant difference with p<0.05.

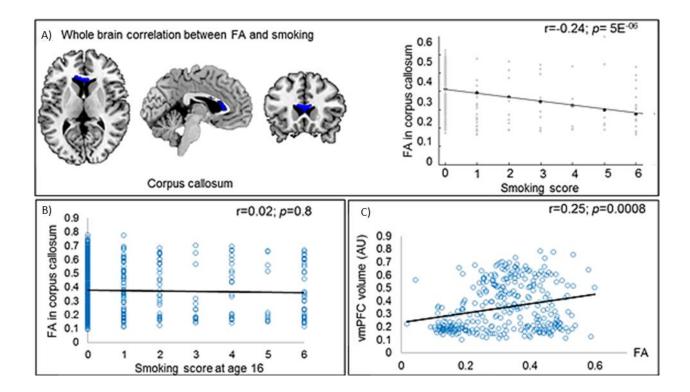
Figure 3. Alcohol use and performance IQ association with vmPFC volume in non-smokers. No significant correlations were observed in any of the analyses with $R^2 \le 0.0043$ and p > 0.1, suggesting that alcohol use and performance IQ are not correlated with vmPFC volume. vmPFC: ventromedial prefrontal cortex.

Figure 4. Smoking status effects on white matter integrity. A) Whole brain rendering of the T map and Pearson's correlation showing significant negative correlation between FA values in the corpus callosum ROI and smoking occasions, with r = -0.24 and $p = 5E^{-06}$. B) The regression between the ventromedial prefrontal cortex (vmPFC) volume and future smoking at age 16 in 531 adolescents who were smoking naïve at age 14 revealed no significant correlation relationship with r = 0.02 and p = 0.8. C) Pearson's correlation test highlighting the significant positive correlation between FA values in the corpus callosum cluster and the vmPFC volume, with r = 0.25 and p = 0.00008. (*): significant difference with p < 0.05.









Supplementary Materials

This document contains supplementary Tables 1 and 2.

Analysis		Smoking score	Parental smoking score	Age	Sex (Females)	Performance IQ	Verbal IQ	PDS	Handedness (right handed)	SES	Alcohol use
Regression	Non-Smokers	0	1.4±0.9	14.5±0.4	48 %	111±14	114±13	3.6±0.7	91.1%	18.5±3.6	2.32±1.4
between GMV and	Smokers	2.5±1.8	1.6 ± 0.4	14.5±0.2	50 %	106±13	110±12	3.7±0.6	87.5%	17.7±4	3.8±1.7
smoking scores	Non-smokers vs smokers		NS	NS	$p_{(\chi 2)} = 0.72$	**	NS	NS	$p_{(\chi 2)} = 0.47$	*	**
Regression	Non-Smokers	0	1.3±0.7	14.5±0.3	48 %	111±14	114±13	3.6±0.7	90%	18.3±3.5	2.32±1.4
between FA and	Smokers	2.6±1.8	1.6±0.5	14.5±0.2	50 %	105±13	111±12	3.7±0.6	87.5%	17.9±4	3.8±1.7
smoking scores	Non-smokers vs smokers		NS	NS	NS $p_{(\chi 2)} = 0.72$	**	NS	NS	NS $p_{(\chi 2)} = 0.47$	NS	**
	Non-Smokers	0	1.3±0.8	14.5±0.4	50%	110±14	112±15	3.6±0.7	89%	18.2±3.3	2.2±1.5
Genetic	Smokers	2.4±1.7	1.5±0.3	14.5±0.3	52%	106±13	109±15	3.6±0.6	87%	17.8±4	3.8±1.8
Genetic	Non-smokers vs smokers		NS	NS	$p_{(\chi 2)} = 0.7$	**	NS	NS	$p_{(\chi 2)} = 0.5$	NS	**

Supplementary Table 1. Demographic information for the participants in regression and genetic analyses. Quantitative measures are expressed as mean \pm standard deviation. NS: non-significant difference; (*): t test p < 0.05; (**): t test p < 0.005. (GMV): Grey Matter Volume; (FA): Fractional Anisotropy; (PDS): Pubertal Development State.

Locus	Gene	HW P value	MAF	Major: Minor alleles	Imputation quality (R ²)	LD measures		Association with smoking		Genotype % AA:GA:GG	
						D'	\mathbb{R}^2	β	p	Non-smokers	Smokers
rs16969968	CHRNA5	0.91	0.35	G:A	0.87	,	0.99	0.037	0.003	12:45:43	13:46:41
rs1051730	CHRNA3	0.9	0.35	G:A	0.92	1		0.038	0.003	12:45:43	13:46:41

Supplementary Table S2. Statistics and frequency of rs16969968 and rs1051730. Measures of Hardy-Weinberg Equilibrium (HW), Minor Allele Frequency (MAF), imputation quality and LD of the two SNPs are reported as observed in the full genotype sample of IMAGEN (1834 subjects). AA, GA and GG are the high-risk, intermediate risk and low-risk genotypes associated with

smoking, respectively. The two SNPs are in tight LD and highly correlated (D'=1; R^2 =0.99). They are significantly associated with smoking (p=0.003) and have similar allele frequencies within non-smokers and smokers.

D': the d' of the linkage equilibrium between the two loci. R^2 , variance explained between the two loci. β : effect size of the association with smoking.

This study shows morphological and functional brain changes associated with low smoking exposure in young adolescent smokers, which may underlie, at least in part, a genetic component associated with smoking. Our findings give insight into a mechanism involving genes, brain structure and brain function underlying why some teens find nicotine especially reinforcing and transition to repeated use leading to increased risk of lifetime dependence.